

COMPARATIVE FINE STRUCTURE OF EGGS OF AUTOGENOUS AND ANAUTOGENOUS *CULEX PIFIENS* (DIPTERA: CULICIDAE)

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Abstract

Culex (Cx.) pipiens is the potential vector of human filariasis in Egypt. However, autogenous *Cx. pipiens* may be less efficient vector of *Wuchereria (W.) bancrofti* in endemic areas of Egypt compared to anautogenous counterparts. In this study, an attempt was made to differentiate eggs of autogenous and anautogenous *Cx. pipiens* using scanning electron micrographs. The results indicated that eggs of both species appear to be similar to great extent in surface morphology. Eggs of both forms are black and elongate-oval. Width is greatest at the anterior end. The posterior end is pointed. The micropylar disc is apparent with distinct edge. Exochorionic bridges are angular. Size of both eggs represented by length and width are comparable. In both eggs, length is greater than width. However, eggs of both forms can be distinguished from each other by the exochorionic bridges being longer and thinner in the autogenous eggs than in the anautogenous eggs.

Key words: Electron Microscope, autogenous, anautogenous, egg, *Culex pipiens*

Introduction

Mosquitoes act as vectors for several diseases including malaria and filaria (Parmanand *et al*, 2008). Despite centuries of control efforts, mosquito-borne diseases are flourishing worldwide (Tolle, 2009). *Culex (Cx.) pipiens* is common mosquito vector so many diseases in urban and suburban areas in the world (Soliman, 1995). It is incriminated as one of the principle vectors of filariasis, Rift Valley fever and other virus-borne diseases. *Culex pipiens* is a polytypic species that exist in autogenous and anautogenous forms (Spielman, 1967). Autogeny, the ability of a mosquito to mature an initial batch of eggs without blood feeding, is an alternative re-productive strategy with important implications for vector-borne disease transmission (Provost-Javier *et al*, 2010). Autogeny is associated with an inherent ability of autogenous mosquitoes to store more nutritional reserves than anautogenous counterparts. Those reserves are the precursors for oogenesis to produce the first egg clutch in autogenous females (Soliman *et al*, 1995). Elevated lipid content before emergence

may play a role of inducing ovarian development in autogenous mosquitoes (Sawabe and Moribayashi, 2000). Larval food must be adequate and abundant, because the quality and quantity of it has a pronounced effect on the autogeny rate (Vinogradova and Karpova, 2006). However, autogenous *Cx. pipiens* may be less efficient vector of *Wuchereria (W.) bancrofti* in endemic areas of Egypt compared to anautogenous counterparts (Hassan *et al*, 1994). The technology required for intervening effectively against vector-borne infection is based on systematic experimental research (Spielman, 2003). Relatively recent mosquito taxonomical studies rely on advanced tools including scanning electron microscopy. The morphometrical and morphological analysis of the egg attributes may be helpful in species confirmation as well as in differentiation between identical forms (Suman *et al*, 2011). The shape of the eggs is generally characteristic of the various genera. The pattern of impressed and raised markings on the outer chorion provides useful taxonomic characters. Suman *et al*. (2009)

suggested that ecological variation may affect the morphometric attributes of the egg of *Cx. quinquefasciatus* strains collected from different geographical areas. On the contrary, no environmental influence on the egg attributes of *Anopheles (An.) darling* was detected (Almeida *et al*, 2014).

Scanning electron microscopy was widely used to describe eggs of *Haemagogus tropicalis* (Alencar *et al*, 2008), *Ochlerotatus albifasciatus* (Santos-Mallet *et al*, 2009), *Aedes (Ae.) scapularis* (Dos Santos-Mallet *et al*, 2010), *Psorophora albigena* (Pacheco *et al*, 2012), *Sabethes cyaneus* (Santos-Mallet *et al*, 2013) and *Georgecraigius fluviatilis* (Sarmiento *et al*, 2014). An ultra-structural key was built for species identification of *An. hyrcanus* group eggs in Thailand based on eggs micrographs (Saeung *et al*, 2014).

The present research dealt with comparative study by scanning electron microscopy of surface morphology and morphometrics of Egyptian autogenous and anautogenous *Cx. pipiens* eggs.

Materials and Methods

Anautogenous *Cx. pipiens* larvae were collected from a drainage canal in Suez Governorate. Autogenous *Cx. pipiens* larvae were collected from cement water basin located in the Garden of Ain Shams University. Larvae were raised in a walk-in insectary under optimum conditions of 27±2°C

and fed on fish food till pupation. Emerged adults were allowed to mate for 24-48 hours. Anautogenous female mosquitoes were offered a blood meal followed by sucrose solution, while autogenous mosquitoes were fed on sucrose solution only. Eggs from either group were allowed to embryonate for 24 hours, and then fixed in bouin solution according to Linley *et al*. (1996). Eggs were then rinsed in three changes of 80% ethanol to remove picric acid, dehydrated through a continuing ethanol series and dried by the critical point method. Eggs were set with a fine artist's brush on stubs coated with carbon, dried, and then coated with gold.

Descriptions: Autogenous form (Figs 1 -2, Tab. 1)

Size: length (mean 494.75±10.76 µm, n=4) is greater than width (mean 98.50±14.15 µm), as indicated in length/width ratio (mean 5.37±0.85). Color: black

Shape: elongate-oval, width greatest at the anterior end, posterior end pointed. Micropylar collar: micropylar disc apparent, edge distinct (Fig.1a, b).

Posterior end: pointed compared to the anterior end (Fig. 1 c, d), boundaries of outer chorionic cell fields angular (Fig. 2 e, f). Exochorionic bridge longer and thinner than anautogenous \ (length 0.61±0.05 µm, width 0.21±0.01 µm).

Table 1: Dimensions of eggs of autogenous (n=4) and anautogenous (n=5) *Culex pipiens*

Form	Length (µm)		Width (µm)		L/W Ratio	
	Mean ± SE	Range	Mean ± SE	Range	Mean ± SE	Range
Autogenous	494.75 ± 10.76	477-523	98.50 ± 14.15	60.7-129	5.37 ± 0.85	4.05-7.86
Anautogenous	480.60 ± 8.49	461-503	88.64 ± 7.03	63.6-103	5.61 ± 0.60	4.57- 7.9

Table 2: Dimensions of exochorionic bridges of autogenous (n=6) & anautogenous (n=8) *Culex pipiens*

Form	Length (µm)		Width (µm)	
	Mean ± SE	Range	Mean ± SE	Range
Autogenous	0.61±0.05	0.47-0.79	0.21±0.01	0.18-0.23
Anautogenous	0.46 ± 0.02	0.40-0.60	0.30 ± 0.02	0.24-0.42

Significantly different P ≤ 0.05

Anautogenous form (Tab.1): Micropylar collar: micropylar disc apparent, Size: length (mean 480.60±8.49 µm, n=5) greater than width (mean 88.64±7.03 µm), a indicated

in length/width ratio (mean 5.61±0.60). Color: Black. Shape: elongate-oval, width greatest at the anterior end, posterior end pointed, edge distinct (Fig. 3 a, b). Posterior end:

pointed compared to the anterior end (Fig. 3 c, d). Exochorionic bridges angular (Fig. 4 e, f): Outer chorionic reticulum: shorter ($P=0.01$) and thicker ($P=0.002$) than autogenous (length $0.46\pm 0.02\mu\text{m}$, width $0.30\pm 0.02\mu\text{m}$).

Discussion

Culex pipiens L. is the most widespread mosquito vector in the temperate regions (Gomes *et al*, 2009). This species consists of two forms, denoted *molestus* and *pipiens* that exhibit important behavioural and physiological differences. *Culex pipiens pipiens* and *Cx. pipiens* Forskål *molestus* differ little morphologically, but their biological features differ significantly. *Culex pipiens* Forskål *molestus* form is autogenous, stenogamous and anthropophilic, whereas *pipiens* form is anautogenous, eurygamous and capable of diapause. Both forms occur in the sympatry almost throughout the whole area of distribution (Vinogradova, 2000). In the U.S.A., *Cx. pipiens* bioforms, *Cx. pipiens* form *pipiens* and *Cx. pipiens* form *molestus*, as well as *Cx. quinquefasciatus*, reside in distinct but overlapping ecological niches and readily hybridize in areas where they coexist (Ciota *et al*, 2013). *Culex molestus* is a phenotypic and physiological variant of *Cx. pipiens* (Harbach, 2012). The sympatric presence of different *Cx. pipiens* biotypes was also confirmed in Portugal, Western Europe (Osório *et al*, 2014).

Culex molestus Forskål was originally described from Egypt as a distinct species in 1775. It was recorded in Europe from the 1920s, first in London, England, and later in many other countries (Mihalyi, 1965; Vinogradova, 2000). In the temperate zone, both forms differ in their habitats. The urban mosquitoes develop throughout the year in underground sites flooded by polluted waters, e.g. in the basements of houses and underground tunnels. In this zone both forms are strongly isolated from one another. Such situation was observed in London, England (Byrne and Nichols, 1999) and in the Upper Rhein Valley in Germany (Becker *et al*, 1999). The degree of isolation between

the two forms decreased in the southern of Europe, the two forms live in sympatry in surface habitats which promotes hybridization between the two species. As a result, populations with intermediate biological characteristics have been described. For instance, Egyptian populations of *Cx. pipiens* are relatively homogenous (Farid *et al*, 1991; Vinogradova and Shaikevich, 2007). Until the relatively recent application of molecular identification tools, identification of *Cx. pipiens* f. *pipiens* and *Cx. pipiens* f. *molestus* relied on expressed ecological characteristics, including autogeny, host preference and stenogamy (Danabalan *et al*, 2012). From the epidemiological point of view, autogenous *Cx. pipiens* may be less efficient vector of *W. bancrofti* in endemic areas of Egypt compared to anautogenous counterparts (Hassan *et al*, 1994).

Similarly, preference of *Cx. molestus* to delay blood feeding until up to day 8 following emergence may reduce their relative importance as a vector of arboviruses (Kasim *et al*, 2012).

The present results indicated that *Cx. pipiens* eggs of autogenous and anautogenous forms appear to be similar to great extent in surface morphology. This result is supported by the fact that there is no limitation for hybridization between autogenous and anautogenous *Cx. pipiens* mosquitoes in Egypt due to the lack of premating and postmating barriers (Nudelman *et al*, 1988; Gad *et al*, 1995). Egyptian populations of *Cx. pipiens* are confirmed to be relatively homogenous (Farid *et al*, 1991; Vinogradova and Shaikevich, 2007). However, the exochorionic bridge served to differentiate the two forms. The exochorionic bridge of autogenous egg was longer and thinner than that of anautogenous egg. Previous studies agreed that outer egg chorion had served to recognize *An. dunhami* from the related *An. trinkae* Faran and *An. nuneztovari* (Lounibos *et al*, 1997) and to differentiate between *Cx. tritaeniorhynchus* and *Cx. quinquefasciatus* (Suman *et al*, 2008) and between *Ae. aegy-*

pti and *Ae. albopictus* (Suman *et al.*, 2011).

Soliman *et al.* (2014) suggested that the two forms autogenous and anautogenous *Ae. (Ochlerotatus) caspius* of Egypt are two distinct species depending on morphometric differences in the ultrastructure micrographs of eggs. In the present study, it can be concluded that the morphometric variations of the egg micrographs of the two Egyptian autogenous and anautogenous *Cx. pipiens* forms are not sufficient to judge that this study were dealing with two distinct species and confirmed the hybridization theory resulting in populations with intermediate biological characteristics (Farid *et al.*, 1991; Gad *et al.*, 1995; Vinogradova and Shaikevich, 2007).

Conclusion

Future study must focus on the recent application of molecular tools to solve the problem of *Cx. pipiens* complex in Egypt.

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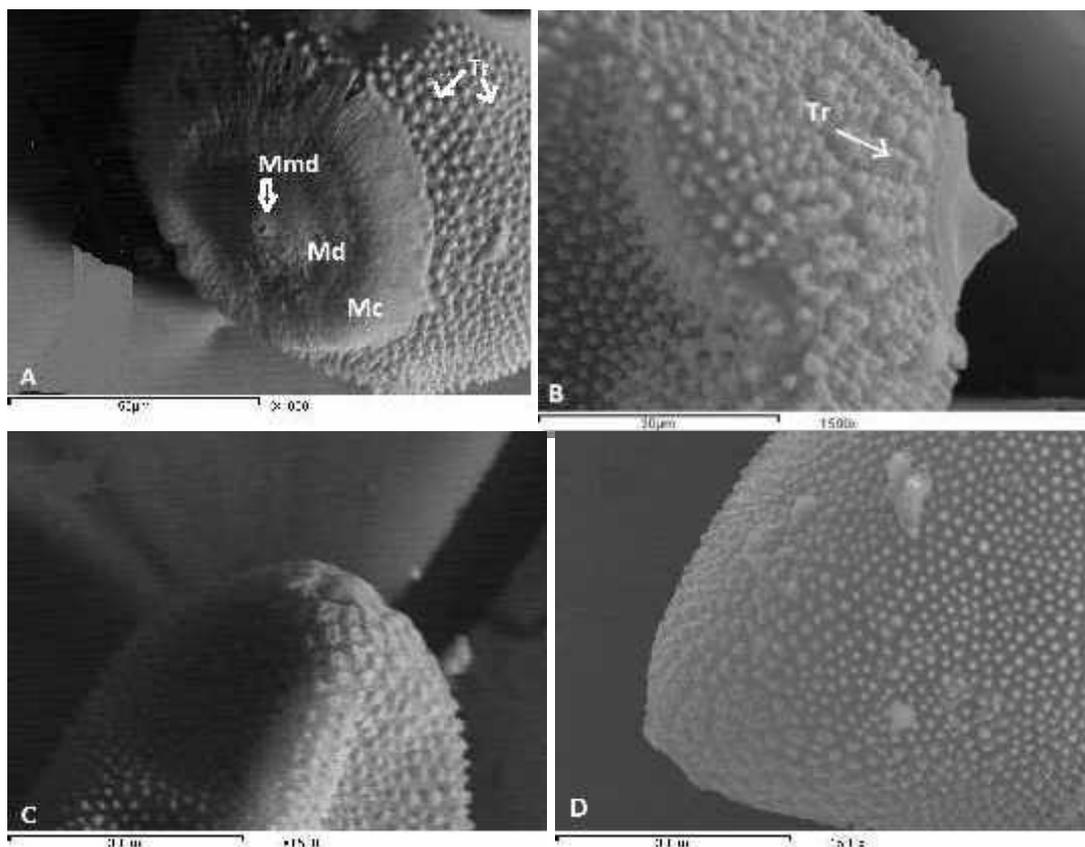


Fig. 1: Scanning electron micrographs of egg surface of autogenous form of *Culex pipiens*. A: anterior end; B: micropylar region; C: posterior end; D: posterior end, lateral view. (Mc: micropylar corolla, Md: micropylar disc, Mmd: micropylar mound, Tr: tubercular rows)

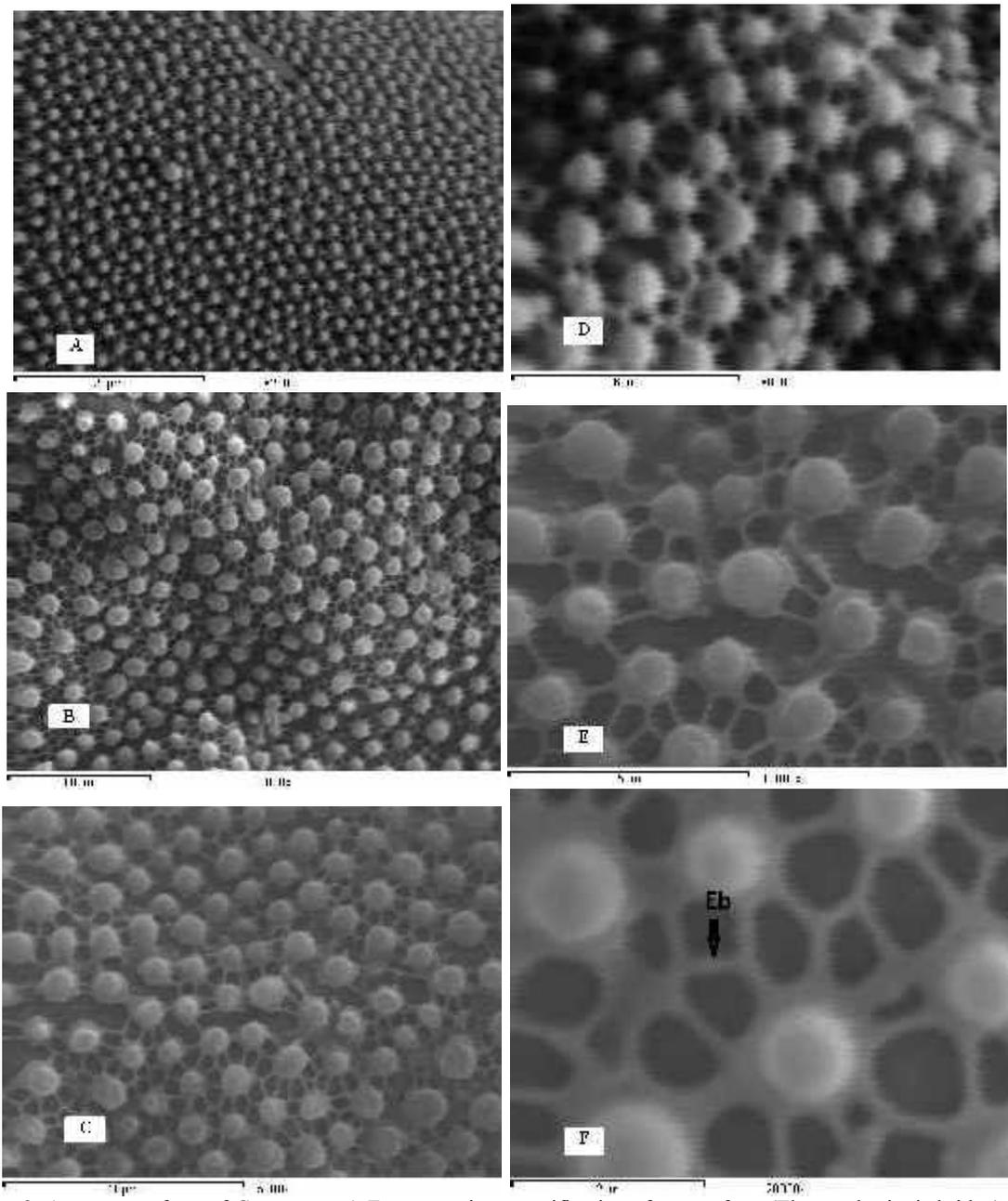


Fig. 2: Autogenous form of *Cx. pipiens*. A-F: progressive magnification of egg surface. (Eb: exochorionic bridge)

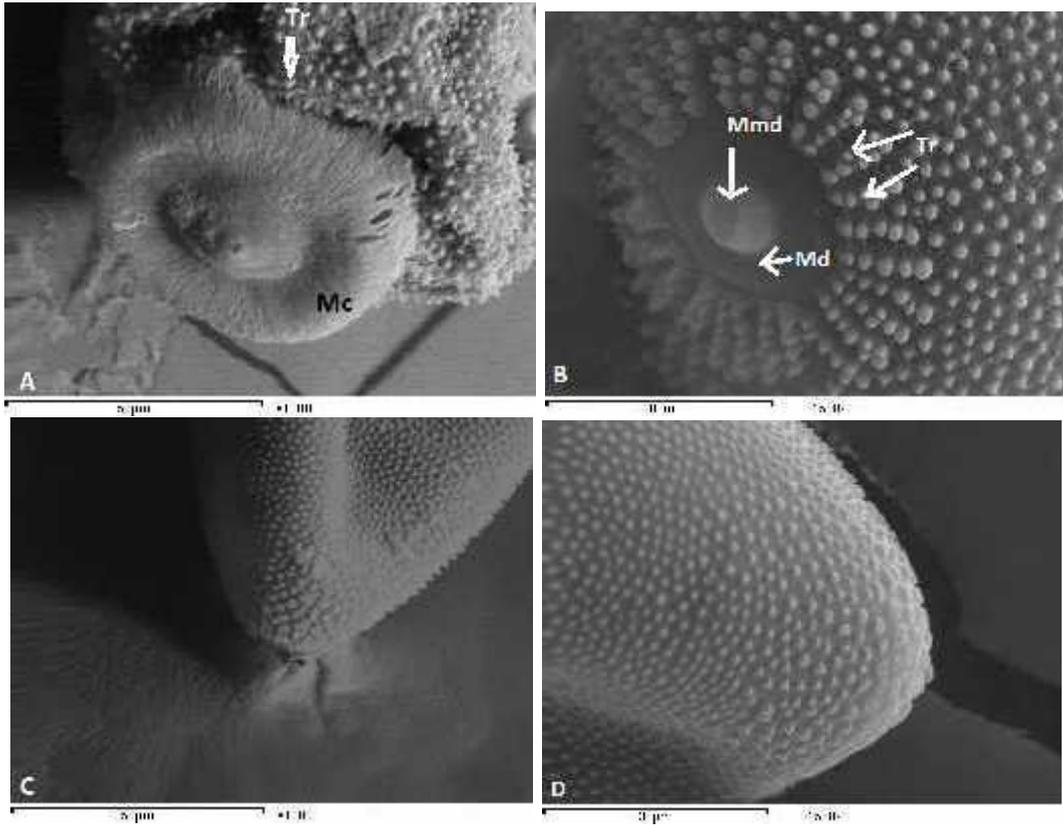


Fig. 3: Anautogenous form of *Culex pipiens*. A: anterior end; B: micropylar disc; C: posterior end; D: posterior end, lateral view. (Mc: micropylar corolla, Md: micropylar disc, Mmd: micropylar mound, Tr: tubercular rows)

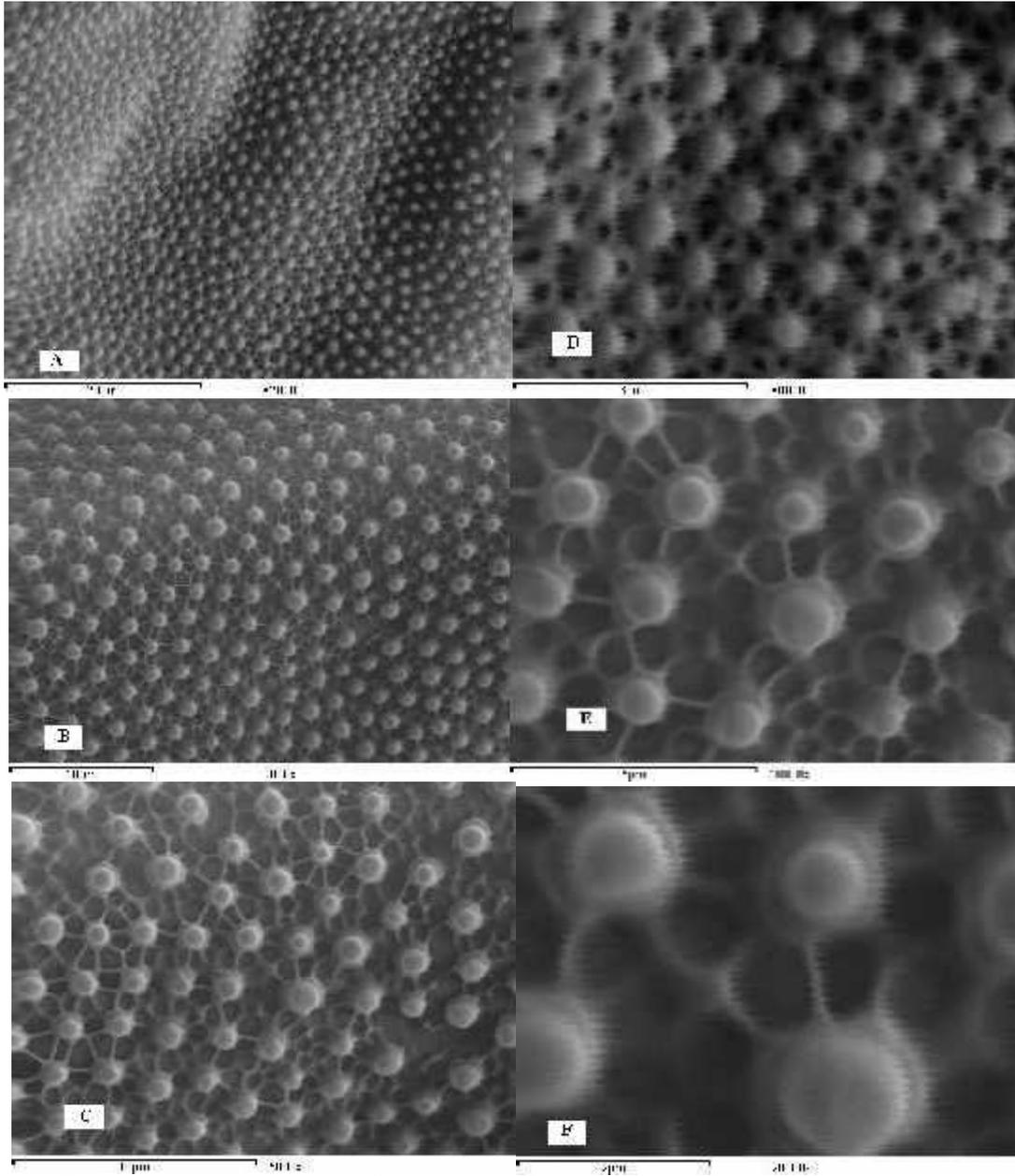


Fig. 4: Anautogenous form of *Cx. pipiens*. A- F: progressive magnification of egg surface.