

**CHARACTERIZATION OF HLA-DRB1 ALLELES POLYMORPHISMS IN
PLASMODIUM FALCIPARUM MALARIA INFECTIONS IN JAZAN, KSA**

By

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Abstract

Malaria is a public health problem in the KSA. It is restricted to the southwestern region, where *P. falciparum* is the most prevalent species. The immune response which affects the clinical outcome of malaria is genetically controlled and is influenced with high degree of HLA genetic polymorphism. The present study aimed at assessing the relationship between HLA-DRB1 alleles and malaria susceptibility pattern moreover its degree of severity in a group of Saudi population in Jazan, KSA. A total of 60 malaria cases (51 uncomplicated and 9 severe malaria cases) and 60 control subjects were enrolled in this study. Blood samples were collected and genotyping of the HLA-DRB1 alleles was performed. The results suggested significant associations of HLA-DRB1* 04 and HLA-DRB1* 11 with susceptibility to malaria ($p = 0.002$ and $p = 0.043$, respectively) and association of HLA-DRB1* 13 with resistance to malaria ($p = 0.032$). The HLA-DRB1* 04 was found to be associated with severe malaria.

The individuals with HLA-DRB1* 04 and HLA-DRB1* 11 were at increased risk to malaria infection and the HLA-DRB1* 04 carriers are more susceptible to severe malaria. While, HLA-DRB1* 13 carriers are more resistant to this infection. Wide investigations for this association may lead to implementation of a successful effective malaria vaccine program.

Key words; Saudi Arabia, *Plasmodium falciparum*, HLA-Drb1 Alleles Polymorphisms

Introduction

Malaria is one of the most prevalent endemic diseases worldwide (Autino *et al*, 2012). It has been a life-threatening illness in many countries for thousands of years, and has long been a challenge to be eradicated all over the world (Bawah and Binka, 2007). Almost half of the total world population is exposed to the risk of contracting malaria (Dalrymple *et al*, 2015). In 2016, an estimation of 216 million cases of malaria associated with 445000 deaths was reported (WHO, 2017).

In Saudi Arabia, about 5% of the national population (1.04 million people) is at risk of infection. Malaria occurs at hypo-to hyper perendemic level where *P. falciparum* was the commonest parasite that accounts for over 87% of malaria cases in Saudi Arabia (Alzahrani *et al*, 2017). However, the implementation of extensive control programs made it highly restricted to the Southwestern region. Jazan that is located in the Southwestern area and bordering Yemen (the most

malarious regional country), estimated to be responsible for most locally acquired malaria cases recorded in Saudi Arabia (Bin Dajem and Al-Qahtani, 2010). More than 70% of total national case burden occurred mainly Jazan and Asir (Snow *et al*, 2013). The average annual incidence of 0.3/10,000 of the population was reported in Jazan through 2010-2014 (El Hassan *et al*, 2015). Meanwhile, the highest percentage of chloroquine resistant *P. falciparum* malaria was reported in Yemen which borders Saudi Arabia (Al-Maktari and Bassiouny, 2003).

Plasmodium spp. leads to diverse clinical manifestations ranging from asymptomatic to severe complicated malaria (Bartoloni and Zammarchi, 2012). This wide spectrum of clinical presentation relay to a large extent on the underlying pathophysiological mechanism that differ from person to person, by presence in the same epidemiological circumstances which indicate different immune reactivity to malaria (Lima-Junior and Pratt-Riccio, 2016). In *P. falciparum*

endemic areas, severe malaria is commonly found in children aged 5 years or less, while gaining partial immunity in older children and adults make the disease less severe. In areas with low endemicity, the age distribution of severe malaria is not well demarcated and may also occur in adult semi-immune persons (Bartoloni and Zammarchi, 2012). Several factors control malaria infection tendency (Belachew, 2018). One of the most important of these factors is the possible relationship between malaria and Human Leukocyte Antigen (HLA) gene variations that lead to a wide variability in immune reactivity (Driss *et al*, 2011). Utilization of HLA genes as a marker in malaria pathogenesis is substantially highlighted playing a crucial role in host-pathogen interaction (Lima-Junior and Pratt-Riccio, 2016). HLA system is divided into three classes: class I: the classical HLA genes (HLA-A, -B & -C), class II: the immune response genes; HLA-DR, -DQ, and -DP, each of them has A & B sub-unit and class III genes are for complement components (C2, C4, factor B), 21-hydroxylase, tumor necrosis factors (TNFs), & other immune variants (Choo, 2007). In Class II, most variability comes from HLA-DRB, with >700 known alleles, but only three HLA-DRA variants are present (Driss *et al*, 2011). The relationship between class I and class II HLA alleles and malaria infection has been widely investigated; interestingly, this relationship was studied for both within-countries level (intra-countries) and between countries level (inter-countries) with more expressive results for between countries level (Garamszegi, 2014). eg. In an Indian case control study, a group of alleles: A3, B27, B49, DRB1*04, and DRB1*0809 showed increased incidence of malaria, while A19, A34, B18, B37, & DQB1*0203 showed decreased incidence. HLA B49 & DRB1*0809 were associated with the complicated severe form. HLA A19, B5 and B13 were protective in high parasite index patients (Shankarkumar *et al*, 2002).

But, in West Africa, some alleles were ass-

ociated with the sever malaria resistance: B*53:01, DQB1*05:01, DRB1*13:02 (Hill *et al*, 1991), or associated with susceptibility to sever malaria: DRB1*04:01 (May *et al*, 1999). In this concern ethnic background must be taken into consideration while developing an ideal malaria vaccine (Shankarkumar *et al*, 2002). Wide investigations for this association may lead to implementation of a successful effective malaria vaccine program (Kwiatkowski, 2005).

This study aimed to assess the relationship between HLA-DRB1 alleles and malaria susceptibility pattern and its degree of severity in a group of Saudi population in Jazan, Kingdom of Saudi Arabia.

Subjects and Methods

Patients and controls: Through a case control study, two groups of subjects were selected 60 subjects each. Malaria cases group were selected from patients referred to Prince Mohamed Bin Naser, Sabia, Abo-Erish and El-Ardaa general hospitals. They were 37 males and 23 females with positive diagnosis for *Plasmodium falciparum* they were selected from September 2015 to September 2018. Malaria diagnosis was based on clinical symptoms, the rapid diagnostic antigen detection OptiMal test (ICT-Amrad, Sydney, Australia) and examination of Giemsa stained thin & thick blood films. Parasitaemia levels were calculated against WBCs and expressed as number of parasites/ μ l. Control group included 60 Saudi healthy individuals age and sex matched to cases without present or past history of malaria.

Malaria was classified into uncomplicated malaria and severe malaria (Bartoloni and Zammarchi, 2012), as follows: a- Uncomplicated malaria: Pyrexia, flu-like illness, myalgia, arthralgia, lassitude, headache, anorexia, vomiting, abdominal pain, lower back pain, tachycardia, and b- Severe malaria: The presence of one or more of the following manifestations is sufficient for a diagnosis of severe malaria: Impaired consciousness/coma, convulsions, hypoglycae-

mia, severe normocytic anaemia, metabolic acidosis, coagulopathy with DIC, hyperparasitaemia >5%, pulmonary oedema, ARDS, acute renal failure, haemoglobinuria, circulatory shock, splenic rupture.

The patients and controls were included in after informed with the aim of the study and an accepted written consent was obtained from each one. The study was approved by the Scientific Research Ethics Committee (No. REC 39/4FS001) Jazan University.

Blood samples collection and DNA extraction: Peripheral blood samples were obtained from patients and controls in EDTA sterile vacutainers. DNA extraction and purification were performed from the whole blood samples using G-spin™ total DNA extraction kit (iNtRON Biotechnology) following the manufacturer's instructions. DNA concentrations were obtained using the NanoDrop One/One^c spectrophotometer (Thermo Fisher Scientific) and content measured between 0.02 & 0.04 μ g/ μ l was used for HLA-DRB1 amplification by PCR.

HLA-DRB1 alleles genotyping: Genotyping of HLA-DRB1 alleles was done using the INNO-LiPA HLA-DRB1 Plus (Fujirebio, Germany GmbH) following the manufacturer's instructions.

The test was based on the reverse hybridization principle and the following steps were involved: Step 1: Amplification of exon 2 of

Table 1: Characteristics of the malaria cases group in relation to malaria severity.

Character	Uncomplicated malaria (n=51)	Sever malaria (n=9)	Total (n=60)
Age group:			
≤ 5 Years	7	4	11
>5-15	6	1	7
15-25	4	0	4
25-35	7	1	8
35-45	9	0	9
45-55	10	1	11
> 55 years	8	2	10
Sex:			
Male	32	5	37
Female	19	4	23
Total	51	9	60

HLA-DRB1 alleles frequencies in patients and controls: 12 alleles were detected in them, the commonest frequent allele in patients was HLA-DRB1 * 04, with frequency

HLA-DRB1 alleles. Step 2: Hybridization and stringent wash with 37 probes immobilized on one INNO-LiPA HLA-DRB1 Plus strip (56°C). Step 3: Color development. Step 4: Interpretation of the probe reactivity pattern using the LiRAS interpretation software.

Statistical analysis: Data were analyzed using SPSS v16 (SPSS Inc, Chicago, USA). Different HLA-DRB1 allele frequencies were compared between pairs of malaria groups to assess association of different alleles with disease susceptibility or resistance by using Chi-square (χ^2) test based on a 2x2 contingency table. P value was considered significant at $P \leq 0.05$.

Results

In the present study 120 Saudi subjects (60 malaria patients and 60 healthy controls) were tested for HLA-DRB1 alleles frequency. The demographic characteristics of malaria cases were shown with a mean age \pm (SD) 33 \pm 9 years (Bartoloni and Zammarchi, 2012), malaria cases were grouped into 51 uncomplicated malaria cases and 9 severe malaria cases.

The most frequent severe malaria cases occurred in age group \leq five years (4 cases) and to a lesser degree for cases more than 55 years (2 cases), malaria infection was prevalent in males more than females (37, 62% vs. 23, 38%).

of 28% and the commonest one in controls was HLA-DRB1 *13 with frequency of 22%, frequency of 3% was the least frequency detected as it was found in patients for allele

HLA-DRB1*08 and in controls for alleles HLA-DRB1*07 and HLA-DRB1*11.

Significant difference was between pa-

tients and control in alleles HLA-DRB1*04, HLA-DRB1*11 and HLA-DRB1*13. P= 0.002, 0.043 & 0.032 respectively (Tab. 2).

Table 2: Distribution of HLA-DRB1 alleles in malaria cases and controls

HLA-DRB1 allele	Malaria cases		Control		Chi square	P value ^a	95% CI	OR
	No.	%	No.	%				
HLA DRB1* 03								
Positive	11	18	6	10	1.581	0.208	4.73 to 20.71	2.020
Negative	49	82	54	90				
HLA DRB1* 04								
Positive	17	28	4	7	9.087	0.002 ^a	7.47 to 34.1	5.534
Negative	43	72	56	93				
HLA DRB1* 07								
Positive	3	5	2	3	0.310	0.577	6.54 to 10.98	1.526
Negative	57	95	58	97				
HLA DRB1* 08								
Positive	2	3	4	7	1.002	0.316	4.92 to 13.62	0.482
Negative	58	97	56	93				
HLA DRB1* 09								
Positive	7	12	9	15	0.229	0.632	9.63 to 15.63	0.748
Negative	53	88	51	85				
HLA DRB1* 10								
Positive	8	13	5	8	0.791	0.373	6.53 to 16.71	1.692
Negative	52	97	55	92				
HLA DRB1* 11								
Positive	9	15	2	3	5.231	0.022 ^a	1.52 to 23.33	5.117
Negative	51	85	58	97				
HLA DRB1* 12								
Positive	6	10	8	13	0.26	0.608	8.96 to 15.02	0.722
Negative	54	90	52	87				
HLA DRB1* 13								
Positive	5	8	13	22	4.573	0.032 ^a	1.06 to 26.83	0.293
Negative	55	92	46	77				
HLA DRB1* 14								
Positive	9	15	7	12	0.229	0.632	9.63 to 15.63	1.336
Negative	51	85	53	88				
HLA DRB1* 15								
Positive	7	12	4	7	0.865	0.352	6.12 to 16.37	1.849
Negative	53	88	56	93				
HLA DRB1* 16								
Positive	5	8	9	15	1.432	0.231	4.86 to 19.03	0.515
Negative	55	92	51	85				

^a Significant at p ≤ 0.05, CI: confidence interval, OR: odds ratio. Only alleles with frequency shown

According to malaria severity (Table 3) one allele HLA-DRB1*04 showed signifi-

cant difference between uncomplicated and sever malaria patients (P = 0.006)

Table 3: Association of HLA-DRB1 alleles with malaria severity.

Malaria severity	Uncomplicated malaria (51)		Severe malaria (9)		P value ^a
HLA-DRB1 allele	No.	%	No.	%	
HLA DRB1* 03					
Positive	8	16	3	33	0.230
Negative	43	84	6	67	
HLA DRB1* 04					
Positive	11	22	6	67	0.006 ^a
Negative	40	78	3	33	
HLA DRB1* 07					
Positive	3	6	0	0	0.454
Negative	48	94	9	100	
HLA DRB1* 08					
Positive	1	2	1	11	0.170
Negative	50	98	8	89	
HLA DRB1* 09					
Positive	5	10	2	22	0.307
Negative	46	90	7	78	
HLA DRB1* 10					
Positive	6	12	2	22	0.422
Negative	45	88	7	78	
HLA DRB1* 11					
Positive	8	16	1	11	0.702
Negative	43	84	8	89	
HLA DRB1* 12					
Positive	4	8	2	22	0.202
Negative	47	92	7	78	
HLA DRB1* 13					
Positive	3	6	2	22	0.113
Negative	48	94	7	78	
HLA DRB1* 14					
Positive	6	12	3	33	0.108
Negative	45	88	6	67	
HLA DRB1* 15					
Positive	6	12	2	22	0.422
Negative	45	88	7	78	
HLA DRB1* 16					
Positive	4	8	1	11	0.767
Negative	47	92	8	89	

^a Significant at $p \leq 0.05$, HLA-DRB1 alleles showed significant difference between male & female for HLA-DRB1* 13 ($P = 0.006$). No significant difference between male and female for both HLA DRB1* 04 & HLA-DRB1* 11.

Table 4: Association between gender and significant HLA DRB1 alleles.

HLA DRB1 significant alleles	Male (n=37)	%	Female (n=23)	%	P value
HLA DRB1* 04					
Positive	9	24	8	35	0.36
Negative	28	76	15	65	
HLA DRB1* 11					
Positive	4	11	5	22	0.25
Negative	33	89	18	78	
HLA DRB1* 13					
Positive	1	3	4	17	0.05 ^a
Negative	36	97	19	83	

^aSignificant at $p \leq 0.05$.

Discussion

The association between specific HLA alleles and susceptibility or resistance to *P. falciparum* malaria was previously investigated in several studies among different ethnic groups. Although several associations have been reported yet, no associations have been confirmed in different populations (May *et al.*, 1999; Driss *et al.*, 2011; Mosaad, 2015). The present study results revealed significant associations between malaria infection and a number of HLA- DRB1 alleles, HLA- DRB1*04 & DRB1*11 were associated with susceptibility to infection while DRB1*13 was associated with resistance. Only HLA DRB1* 04 allele was found to be associated with severe malaria.

Studies in other ethnic groups showed different allelic distributions and associations between HLA and malaria. Results from Gabon and Ghana suggested that HLA-DRB1*04 was associated both with susceptibility to malaria in general and an increased risk of severe malaria (Osafo-Addo *et al.*, 2008; May *et al.*, 1999). While in Senegal, The most frequent HLA-DR alleles found in malaria patients were: DR52, DR13, DR10, DR53, DR3 & DR18. A significant difference occurred between patients with severe malaria and the control group of the following alleles: DR3, DR10, DR13. HLA-DR3 was considered as the major marker associated to severe malaria (Ndiaye *et al.*, 1998). In Mumbai, western India, the frequency of DRB1*04, & DRB1*0809 were significantly increased in patients with malaria compared to controls. DRB1*0809 was found to be positively associated with the complicated severe malaria (Shankarkumar *et al.*, 2002). Another study in Thai malaria patients showed a significant difference in distribution of HLA-B46, -B56 & DRB1*10:01 between malaria groups; mild malaria, non-cerebral severe malaria and cerebral malaria groups. DRB1*10:01 was significantly increased in mild malaria group compared to non-cerebral severe malaria (Hannantachai *et al.*, 2005). On the other hand,

Carpenter *et al.* (2009) in Tanzania studied the genetics of susceptibility to malaria related phenotypes population and concluded weak associations between HLA-DRB1*04 and HLA-DRB1*10 with parasite density.

In the present study, HLA-DRB1*11 was associated with malaria infection. Although it was not reported previously to be associated with malaria, it was reported to be associated with other diseases. Associations were reported between HLA-DRB1*11 and persistence of hepatitis B virus infection (Thio *et al.*, 1999); Mite sensitive asthma (Lara-Marquez *et al.*, 1999); juvenile rheumatoid arthritis (Garavito *et al.*, 2004) and systemic sclerosis (Kuwana *et al.*, 1999).

In the present study, HLA-DRB1*13 was associated with reduced susceptibility to malarial infection as the difference between patients and control was statistically significant. In agreement with this result, Hill *et al.* (1991) found that HLA-DRB1*1302, B*53:01 & DQB1*05:01 were associated with resistance to severe malaria in a case-control study of severe malaria in children in Gambia. However, May *et al.* (1999) failed to observe protective effect of DRB1*1302 in a group of patients in Gabon. While in Western Kenya, an association between HLA DRB1*0101 and resistance to severe malaria was reported (Hill, 1998). Others suggested that HLA DQB1*0203 was associated with decreased risk of malaria infection (Shankarkumar *et al.*, 2002).

On the other hand, Lyke *et al.* (2011) observed that the allele frequency distribution of HLA-A differed significantly between patients with cerebral and uncomplicated malaria, whereas there was no significant difference found for HLA-B, -C nor -DRB1 in a population of Malian children. Moreover, in Gambia no association was between HLA & frequency of clinical episodes of malaria (Bennett *et al.*, 1993). The differences observed in the present study results compared to other studies could be due, in part, to variation in allele frequencies and the geographic variation in the genetic

makeup of different ethnic groups (Torcia *et al.*, 2008; Ockenhouse *et al.*, 2006). This genetic diversity has been shown to contribute to differences in immune response to diseases or disease outcomes (Ockenhouse *et al.*, 2006). Also, numerous factors are thought to be responsible for the emergence of different dominant HLA antigens in different endemic populations. Such factors include micro heterogeneity in parasite species, wide genetic variability in parasitic antigens, transmission intensity fluctuations, different polymorphisms of red cell antigens, and selection pressure that have been exerted by diversity in the HLA system (Hedrick, 2011).

The mechanism associating HLA-DRB1 alleles with susceptibility or resistance to malaria was studied by several researchers. In this respect, the antigen presentation of MHC class I proteins, by triggering cytotoxic T cells against intracellular parasites, may play an important role during the liver-stage infection, while class II molecules can mediate the clearance of parasitized erythrocytes from bloodstream through the stimulation of helper T cells by providing help for B cell differentiation and secretion of antibodies (Wijayalath *et al.*, 2014). The CD4+ T cells also restrict growth of erythrocyte *Plasmodium* parasites through cytokine secretion and macrophage activation (Tsuiji, 2010). Nardin *et al.* (2000) found higher anti-sporozoite antibodies titers associated with HLA-DR5 (DRB1*1101) plus DRB1*0401 and DQB1*0603 in a clinical trial of Multiple Antigens Peptides (MAP) vaccine. Storti-Melo *et al.* (2012) found an association between HLA-DR3 & HLA-DR5 alleles and lack of antibody response to CSP (Circum Sporozoite Protein) amino terminal, as well as an association between HLA-DR3 and the highest antibody response to MSP1 (Merozoite Surface Protein 1) in Brazilian population exposed to malaria. Lima-Junior *et al.* (2012) found high frequency of responders to carboxy-terminal (CT) and aminoterminal (NT) regions of MSP-3 in HLA-DRB1*04 carriers. Presence of HLA-DRB1

*04 was positively associated with the IgG immune response against the amino-terminal domain (NT) and the C-terminal blocks of tandem repeats (RII & RIRII) of MSP-9. Wijayalath *et al.* (2014) proved that HLA-DR0402, HLA-DQ6, HLA-DQ8, and to less extent, HLA-DR0301 molecules sufficed for supporting antibody responses and self-curing of *Plasmodium yoelii* strain 17XNL malaria infection, while HLA-DR0401 molecules failed to do so.

For the current study, HLA-DRB1*13 that was associated with reduced susceptibility to malaria, was found more prevalent in females than males in a significant difference. Akanbi *et al.* (2010) in Nigeria concluded that the mean malaria parasite density was higher in male than in female patients. Sex differences in immune status and immune response to parasites were widely studied in vertebrates (Klein *et al.*, 2004) It was reported that females had higher immune responses than males conspecifics particularly, innate immune responses, antibody-mediated responses, and cellular responses (Roberts *et al.*, 2001). Also, the antigen-presenting cells (APC) of female mice were more efficient at presenting peptides compared to male mice (Weinstein *et al.*, 1984). Also, Klein *et al.* (2008) in vivo suggested that physiological levels of estrogen, rather than progesterone, enhanced immunity and, possibly, protected females from disease symptoms during malaria infection. Also another study demonstrated that protection by vaccines developed against *P. chabaudi* in mice was greater for females than males and that elevated testosterone concentrations reduced the efficacy of vaccines against *P. chabaudi* (Wunderlich *et al.*, 1993). Exposure of adult female mice to testosterone reduced the antibody production, decreased major histocompatibility complex (MHC) class II cells in spleen, and increased the CD8+ T-cells (Benten *et al.*, 1997). Legorreta-Herrera *et al.* (2015) reported that in female mice, sex hormones have anti-inflammatory properties and sex hormones levels

affect the immune response and must be considered when designing malaria vaccines.

Conclusion

The individuals with HLA DRB1* 04 and HLA DRB1* 11 are at increased risk to malaria infection moreover HLA DRB1* 04 carriers are more susceptible to severe malaria. While, HLA DRB1* 13 carriers are more resistant to this infection. The disclosure of genetic markers associated with various malaria phenotypes will help clarify the pathophysiology of malaria and empower development of interventions and cures.

Author contribution: Wafaa M. Zaki proposed the research idea, the study design, contribution in sample collection, laboratory procedures, and prepared the final version for submission. Hanan Z. Ryan shared in the study design, sample collection, and revised the manuscript. Aymen M. Madkhaly was responsible for acquisition, analysis and interpretation of resulting data, Khawlaa M. Madkhaly contributed in sample collection and laboratory procedures, all authors revised manuscript and approved it.

Conflict of interest: The authors declare having no conflict of interest.

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