

TOLERANCE OF *HETERACRIS LITTORALIS* (ORTHOPTERA: ACRIDIDAE) TO THE TOXIC PLANT, *NERIUM OLEANDER* (APOCYNACEA)

By

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Abstract

Nerium oleander (Apocynaceae) is a highly toxic plant that has never been reported to be selected as a food or even as a resting place by any acridid species. The study tested the competence of *Heteracris littoralis* (Orthoptera: Acrididae) to tolerate and survive on this plant for considerable periods, and to resume normal capacity of development and reproduction when favourable food plant became available. Nymphal survival and development were followed by monitoring three cohorts of freshly hatched hoppers. The first cohort fed oleander leaves; the second one (control) fed clover; while the third cohort fed oleander leaves for 35 days, and then switched to clover. Age specific survivorship and reproductive parameters of the adults were followed by monitoring three cohorts of newly fledged adults fed on the same patterns. Moulting was suspended in nymphs fed oleander; when food plant was switched to clover, the insects resumed normal development and reached the adult stage. Adult cohort reared on *N. oleander* exhibited a normal age specific survivorship curve, but ovarian development was suspended. Suspension of ovarian development by *N. oleander* was accompanied by very low indices of food consumption and conversion of digested food into body substance. But, after switching to normal food, clover, the insects resumed normal ovarian development with significant prolonged longevity, and the fecundity was approximately equal to the same parameter in the control cohort. This reflects a strong evolutionary biotic potential against the selective power of plant defense.

Key words: Toxic plant, Development, Reproduction, Plant defense, Insect tolerance

Introduction

The Mediterranean Splendid Grasshopper, *Heteracris littoralis* (Rambur) (Orthoptera: Acrididae) is distributed through the Cape Verde Islands across northern Africa to India (Grunshaw, 1991). In Europe, it occurs on the Canary Islands, at the coast of the Iberian Peninsula, in Greece and in Cyprus (Heller, 2013); this grasshopper is a non-graminivorous. Hoppers and adults possess almost the same food taste. Out of several plant species few species including clover, cotton, and bean were found to be favorable as pure diets for survival, development, and reproduction of this grasshopper, with little differences in the growth rates of the hoppers (Ibrahim, 1980). It is generally accepted that secondary plant metabolites are considered as plant defense mechanism against herbivores. Despite the selective force in-

involved in the evolution of plant secondary compounds, they act as feeding deterrents, and can be considered as major determinants of host range of phytophagous insects (Bernays and Chapman, 2000).

Oleander, (*Nerium oleander*: Apocynaceae), is an evergreen urbanite shrub, indigenous to the Indo- Pakistan subcontinent; this plant is widely distributed in the Mediterranean region and subtropical Asia where it is commonly used for ornamental purposes. In Egypt, there are three varieties of *N. oleander*, which can be easily distinguished by the color of the flower (red, pink & white flowered varieties). Longford and Boor (1996) reported that all parts of the plant are toxic to humans, vertebrates and certain insects; while Siddiqui *et al.* (1997) cited that several parts of the plant are therapeutically effective in the treatment of some diseases. Re-

views of the chemistry, pharmacological significance and toxicity of *N. oleander* have been given (Zibbu and Batra, 2010; Hase *et al.*, 2017). Despite its toxicity and pharmacological activity, *N. oleander* supports a complete terrestrial food web of more than 25 arthropod species, representing 20 families, the majority were insects belonged to different feeding guilds and different orders including Lepidoptera, Diptera, Coleoptera, Hymenoptera, Orthoptera, Homoptera, and Neuroptera (El-Shazly, 2002). However, the plant is protected from several herbivores by its defensive secondary metabolites, e.g. *Spodoptera litura* (Rao and Chitra, 2000), *Schistocerca gregaria* (Bagar *et al.*, 2015).

Ecologically, plant as a food resource, is a density-dependent environmental biotic factor; which can be depleted if the population's needs exceeds the available quantities. The unfavourable or toxic plant can be considered as a selective power against insects, if they are forced to devour it confronted with hunger when a favorable plant food resource could not be found. It has been proven that *N. oleander* inhibited moulting in the 4th & 5th instar nymphs of *Schistocerca gregaria* (Bagar *et al.*, 2013); likewise *Cyperusiria* showed pronounced morphogenetic effects on the nymphal development of the grasshopper *Melanopluss anguinipes*, and ovaries dissected from females fed *C. iria* were markedly underdeveloped (Toong *et al.*, 1988). However, no attempt has been made to follow the development and survival if a toxic or unfavourable plant fed to the insect is switched to a favorable one. Indeed, this is approached in the present work. The competence of *H. littoralis* was tested to tolerate and survive for considerable periods on *N. oleander*, as a restricted diet; and the capability of the insect to resume normal capacity of development and reproduction, when a favourable food plant became available.

The study aimed to: a) to test possibility of *H. littoralis* nymphs and adults to survive on

a restricted diet of *N. oleander*, b) to estimate nutritional indices of newly fledged adults fed restricted diet of *N. oleander* compared to favorable food plant (clover), and c) to test the competence of the insect to resume nymphal development and adult reproductive capacity when supplied with only *N. oleander* for considerable periods, before being switched to a favorable food plant.

Materials and Methods

Laboratory culture: Adults and nymphs of *H. littoralis* (Rambur, 1838) were collected from the experimental research farm affiliated to the Faculty of Agriculture, Cairo University, in April 2017. Once in the laboratory, a stock culture was maintained as described by Henry (1985). Individuals of both sexes were placed in wire-screened, wood cages (20×20×30cm) in a rearing room under controlled conditions (28°C±2, 12L: 12D, 65%±5 RH).

The insects were fed with thoroughly washed, fresh leaves of clover, *Trifolium alexandrinum*. The cages were supplied with a substrate for oviposition that consisted of a plastic container (15cm deep) filled with the sterilized moist sand. Mating and thermoregulation were stimulated with 75W bulbs suspended 15 cm above each cage. Egg-pods were incubated *in situ* under the same controlled conditions.

Nymphal survival and development were followed by monitoring three cohorts, each of 60-71 freshly hatched individuals. The control cohort fed favorable food (clover). The oleander cohort was raised on restricted diet of oleander leaves to test the tolerance and survival on oleander as a restricted diet; while the third cohort was fed oleander leaves for 35 days, and then switched to normal food (clover) to test the ability of the insect to resume normal development when favorable food became available. The survival and development were monitored daily until the last survivor moulted to adult; while the oleander-fed cohort was monitored until the last survivor died (because oleander leaves inhibited moulting to adult). Each co-

hort was divided into three groups, kept under controlled conditions of 30°C, relative humidity of 65%±5 and a photoperiod of 12L:12D.

The rate of development after switching the food plant from clover to *N. oleander*, and *vice versa*, was observed by raising two groups of newly hatched nymphs ($N=30$) on restricted diets of either clover or oleander for 23 days, and then the food was altered for 7 days. The insects were weighed and their lengths were measured at suitable intervals.

Nutritional indices of adults: Nutritional indices were determined by gravimetric method on dry weight basis for freshly emerged adults fed oleander leaves compared to control group, fed clover leaves, for 14 days (average preoviposition period of control group). Three replicates, 10 individuals each, were used for both experimental and control groups. The initial fresh food and feces remaining were weighed daily. The quantity of food ingested was determined by subtracting the diet remaining at the end of each experiment from the total weight of fresh diet supplied. To obtain the percentage of dry weight of the food, feces, and insects, 20 specimens for each were weighed, oven-dried (48 h at 60°C), and subsequently reweighed. In this study, “natural losses” including biomass changes in substances other than water (carbon dioxide, volatile materials, microbial decay products, etc.) were not measured (Hemati *et al.*, 2012). The indices were calculated according to the formulae (Wa-ldbauer, 1968): Consumption index (CI) = E/A , Approximate digestibility (AD) = $E-F/E$, Efficiency of conversion of ingested food (ECI) = P/E , Efficiency of conversion of digested food (ECD) = $P/E-F$, Growth rate (GR) = $P/A * T$. A = mean dry weight of insect over unit time, E = dry weight of food consumed, F = dry weight of feces produced, P = dry weight gained by insect, and T = duration of feeding period.

Adult lifespan and reproductive biology:

Age-specific survivorship in the adult stage was followed by monitoring three cohorts, each of 50-60 individuals of freshly fledged adults. One of the cohorts was raised on restricted diet of oleander leaves. The 2nd (control cohort) was raised on restricted diet of clover; while the 3rd cohort fed oleander leaves for 45 days, and then switched to normal food (clover). Because this experiment started from adult emergence, cohort survivorship was recorded on normal food plant (clover) before adult emergence; based on nymphal records, survivorship (lx , proportion of individuals surviving from birth to age x) of each cohort was recorded until the death of last survival in the three cohorts.

Insects of each cohort were segregated in pairs, kept in suitable cages supplied with an ovipositional pot. Reproductive parameters of the cohorts including preoviposition period, longevity, number of egg pods per female, and number of eggs per egg pod were determined. The insects were placed pairwise in wire-screened, aluminium cages (20x20x30 cm) in a rearing room under controlled conditions of 30°C, relative humidity of 65%± 5 and a photoperiod of 12L:12D. The insects were fed with thoroughly washed, fresh leaves of the desired plant.

Results

Heteracris littoralis nymphs of the oleander fed cohort could not reach the adult stage, but survived for considerable periods ranged between 17 and 81 days with an average of 48.25 ± 2.7 days. In control cohort, the normal progressive nymphal development was achieved and the adult stage was reached through 5-7 instars as previously reported for this species (Ibrahim 1980). The nymphal duration ranged between 28 and 38 days, with an average of 28.42 ± 2.21 days and 88.13% survival. In the 3rd cohort, nymphs tolerated feeding on *N. oleander* without moulting for 35 days. After being switched to normal diet, clover, they started moulting through the successive instars, to reach the adult stage in 53 to 63 days (including 35 days feeding on oleander), with

an average of 59.5 ± 2.2 days and 77.97% survival.

The progressive increase in live weights and body lengths of *H. littoralis* male nymphs fed oleander or clover were determined for 30 days. No significance ($P > 0.05$) was recorded between weights and lengths of *H. littoralis* cohorts fed on both diets for the first 10 days, then the difference in the average life weight (Fig. 1) and body length (Fig. 2) increased significantly ($P < 0.05$) in the control cohort.

Two groups of newly hatched nymphs were fed on restricted diets of clover and oleander for 23 days, and then food plant was switched for both groups (Figs. 3 & 4). In the control group, the weight gain of nymphs increased significantly ($P < 0.001$) after 10 days of feeding on clover. After switching the diet to oleander, the weight decreased insignificantly ($P > 0.05$). On the other hand, the weight of nymphs fed oleander for 23 days increased insignificantly ($P > 0.05$) until switching the diet to clover.

Then, the weight of nymphs was not only increased significantly ($P < 0.001$) in one week, but also the nymphs moulted to the next instar (Fig. 3). This was also applied to the body length of the two groups, where the length of the nymphs was significantly ($P < 0.001$) increased twice in one week after switching the diet from the *Nerium* to clover (Fig. 4).

Nutritional indices of *H. littoralis* raised on restricted diet of oleander, compared to control group (raised on clover), were determined for freshly emerged adults for 14 days (the average preoviposition period of the control group). ECI and ECD values, of *H. littoralis* reared on *N. oleander* were significantly lower than those reared on clover (Table 1). ECI is a general index of an insect's ability to use the food consumed for growth and development, while ECD is an index of the efficiency of conversion of digested food into body substance. The consequences of the low ECI and ECD are the week growth rate (GR), while the relatively

high AD on *N. oleander* (Tab. 1) could be a mechanism where-by the insect body compensates the extremely low value of CI. There were extreme differences in the life weights and body size between males and females of *H. littoralis*. The average life weights of freshly fledged adults are 272.10 ± 7.08 and 601 ± 3.98 mg for males and females, respectively; while average lengths of the two sexes were 23.60 ± 0.38 and 37.8 ± 0.56 mm, for males and females, respectively. During the period of sexual maturation, the females raised on normal diet, clover, increased progressively in weight, due to ovarian maturation and the formation of egg pods, to reach 1.33 ± 0.04 mg just before oviposition, while females raised on oleander have shown a slight increase in their live weights during the same period due to the suspension ovarian development (Fig. 5).

The main reproductive parameters of experimental and control cohorts were given (Tab. 2). Adult cohort fed on *N. oleander* from adult emergence, tolerated the toxic effect of the plant and exhibited fairly long lifespans compared to the control cohort (Tab. 2). Meanwhile, the cohort fed as adult on *N. oleander* for 45 days, and then switched to clover, exhibited prolonged lifespans compared to control cohort. The mean longevity of adult males and females fed oleander for 45 days, before being switched to clover, were 1.4 and 1.68 folds more than control male and female groups, respectively. Survivorship curve of the cohort fed on restricted diet of oleander was more or less similar to control cohort, where the mortality was concentrated in the final weeks of adulthood. However, survivorship curve of the cohort fed *N. oleander* for 45 days, and switched to clover showed a more regular death rate (Fig. 6).

Females fed on clover showed well developed ovary (Fig. 7A). Average length of the ovary just before oviposition was 28.9 ± 2.42 mm and full number of ovarioles was 51.3 ± 4.27 . By day 15 after emergence, all females

have already laid their first egg-pod. On the other hand, those fed *Nerium* leaves failed to develop ovaries. The size of ovaries remained small as the newly emerged female (Fig. 7B). These females were also unable to oviposit even after a long period (~45 days). The average number of ovarioles in such females was 25.2 ± 2.57 (Fig. 7C). The two exceptions were observed in two females ($N=30$ individuals), where one egg pod was laid by each of them after 53 & 61 days of their adult lives, respectively. However, the egg pods were unfertile and didn't hatch.

The size of eggs produced in such ovaries was relatively small as compared with normal eggs (Fig. 7D & E). In the cohort fed *N. oleander* for 45 days, sexual maturation and oviposition were suspended until the insects were provided with the favorable food, clover, and then the insects resumed normal development and started to lay eggs within 4 to 8 days; the average number of egg pods per female and the average fecundity were almost the same as the control cohort.

The details were given in tables (1 & 2) and figures (1, 2, 3, 4, 5, 6 & 7).

Table1: Nutritional indices for grasshopper *H. littoralis* adults fed on *N. oleander* and *T. alexandrinum*.

Indices	<i>Nerium</i> -fed adults		<i>Trifolium</i> -fed adults	
	Female	Male	Female	Male
CI	1.957 ± 0.433	0.627 ± 0.041	$12.615 \pm 2.074^*$	$4.854 \pm 1.12^{*m}$
GR	0.0018 ± 0.0001	0.0045 ± 0.0001	$0.404 \pm 0.094^*$	$0.128 \pm 0.007^{*m}$
AD	69.746 ± 7.3168	27.56 ± 3.589	$80.54 \pm 2.57^*$	$25.232 \pm 3.356^{*m}$
ECI	0.5549 ± 0.1132	0.162 ± 0.086	$3.1765 \pm 0.57^*$	$1.059 \pm 0.053^{*m}$
ECD	0.876 ± 0.2513	0.129 ± 0.028	$3.951 \pm 0.98^*$	$1.009 \pm 0.281^{*m}$

* Significant between females at $P \leq 0.05$, *m Significant between males at $P \leq 0.05$.

Table 2: Some biological parameters of *H. littoralis* cohorts fed on restricted diets of clover (control cohort, C1), *N. oleander* (C2) and *N. oleander* switched to clover after 45days (C3).

Parameter tested	C1	C2	C3
Preoviposition period (days)	14.17 ± 2.23 (10-17)	-	50.25 ± 1.64 (49-53)
Male longevity (days)	73.08 ± 6.9 (57-85)	62.12 ± 6.89 (26-76)	102.48 ± 3.44 (48-142)
Female longevity (days)	76.69 ± 5.67 (61-95)	66.64 ± 5.04 (24-77)	129.02 ± 2.36 (37-154)
No. of egg pods/ female	9.75 ± 1.23 (8-11)	-	8.58 ± 1.19 (6-11)
No. of eggs/ egg pod	50.78 ± 7.53^a	-	48.67 ± 5.09^a

Discussion

Tolerance of *H. littoralis* nymphs and adults to restricted diet of *N. oleander* had no apparent influence on their ability to survive and develop when favorable food plant became available. The insects suspended their moulting in the nymphal stage and their ovarian development in the adult stage until favorable food plant became available. Nutritional indices of *H. littoralis* on oleander, as a restricted diet, were estimated during a physiologically defined period, the preoviposition period. During this period the insect exhibits high growth rates and consumes fairly large amounts of food plant under natural conditions. Nathan *et al.* (2005) pointed out that ECD is an index of the efficiency of conversion of digested food into body substance. The slow growth rate (GR) of *H. littoralis* on *N. oleander* could be attributed to

the weak efficiency of conversion of ingested (ECI) and digested (ECD) food into body substance. Analysis of nutritional indices can explain the behavioral and physiological basis of insect's response to host plants (Lazarevic and Peric-Mataruga, 2003).

Heteracris littoralis, like all other acridid species, is not adapted to utilize *N. oleander*; this plant has never been reported to be selected as a food or even as a resting place by any acridid species. In the present study, the insects consumed few amounts of oleander to sustain them alive, instead of rejecting the plant and die. An important fitness indicator of insect population dynamics is body weight (Liu *et al.*, 2004). The low CI and the extremely weak GR of *H. littoralis* on *N. oleander* indicated that, the rate of intake relative to the mean life weight during the feeding period was consistent on this plant

during the feeding period. Meanwhile, the insects suspended their ovarian development. Suspension of ovarian development may be due to the presence of some secondary phytochemicals. Hemati *et al.* (2012) pointed out that Lower fitness of the pod borer, *Helicoverpa armigera* on some host plants may be due to the presence of some secondary phytochemicals in these host plants, or absence of primary nutrients necessary for *growth* and development.

The level of nutrition and the timing of nutritional changes have marked effects on the reproductive output of the lubber grasshopper (*Romalea microptera*) (Moehrlin and Juliano, 1998). Inhibition of moulting and suspension of ovarian development in *H. littoralis* may be attributed to the interference of secondary plant metabolites with the endocrine system of the insect. Some secondary plant metabolites have shown to interfere with the insect's endocrine system as protection techniques against herbivorous insects (Rosenthal and Janzen, 1979). These compounds either act as insect JH analogues (JHAs), or interfere with JH biosynthesis. Previously, Balsam fir juvabione was found to inhibit the metamorphosis of the linden bug, *Pyrrhocoris apterus*, suggesting that plants could use JHAs as defensive secondary metabolites against the insect herbivores (Bowers *et al.*, 1966; Sláma and Williams, 1966). On the other hand, Toong *et al.* (1988) reported the isolation of JH III from the Malaysian plant *Cyperusiria* (Fa. Cyperaceae) along with the closely related compound, methyl (2*E*, 6*E*)-farnesoate; the authors found that ovaries dissected from the grasshopper (*Melanoplus anguinipes*) fed this plant were markedly underdeveloped compared to those of normal females, and considered the isolated compounds as defensive mechanism against insects. In *Locusta migratoria*, JH both primed the fat body and might stimulate production of vitellogenin from the hemolymph (Wyatt and Davey, 1996). Borst *et al.* (2000) suspected that the higher levels of JH III observed at day 18,

during the oviposition cycle of *R. microptera*, stimulated vitellogenin uptake by the oocyte as a rapid rise in the oviposition index occurred. Five diterpene secondary metabolites, with juvenile hormone antagonist (JHANs), were isolated from two plants, *Linderaery throcarpa* and *Solidago serotina*, which represents an innate resistance mechanism of plants against insect herbivores (Lee *et al.*, 2015). Authors hypothesized that JHANs, rather than JHAs, play a significant role in plant defense against insect herbivores. The drastic effect of *N. oleander* on the reproductive power of the *H. littoralis* might be attributed to the existence of strong prohibiting chemical(s) that suppressed egg formation in ovaries. Perhaps *Nerium oleander* blocked the process of vitellogenesis which is controlled by juvenile hormone (Bagari *et al.*, 2015).

Herbivores counter adaptations to allelochemical plant defenses have already been reported via several adaptive mechanisms which involve various behavioral and anatomical adaptations. A possible behavioral mechanism, involved in avoiding *N. oleander* toxic metabolites, might be that phloem feeders avoid exposure to toxic plant secondary metabolites present in mesophyll (El-Shazly *et al.*, 2000). In the present investigation, *H. littoralis* exhibited a strong biotic potential, and was able to break through the defensive barriers of *N. oleander*. Adults reared on this toxic plant, not only exhibited a normal age specific survivorship curve, but they also suspended ovarian development until the food plant was switched to palatable and favorable diet, clover, and then, the insects resumed normal ovarian development with significant prolonged longevity as if, the insect was in a state of dormancy.

These results agreed with Bernays and Chapman (2000) who cited that, polyphagous grasshoppers have a relatively low level of sensitivity to the taste of many secondary compounds, and, if they did respond to the taste, have the capacity to habituate. This gives time for the induction of detoxifying

enzymes so that unpalatable, but potentially nutritious, plants may be eaten safely. The present data showed that physiological and biochemical basis of *H. littoralis* tolerance to the defensive metabolites of *N. oleander* requires more investigations.

Conclusion

Generally speaking, the grasshoppers eat large quantities of foliage both as adults and during their development, and can be serious pests of arid land and prairies. The pasture, grain, forage, vegetable and other crops can be affected. Grasshoppers can best be prevented from becoming pests by manipulating their environment. Shade provided by trees will discourage them and they may be prevented from moving onto the developing crops by removing coarse vegetation from fallow land and field margins and discouraging thick growth beside ditches and on roadside verges.

Undoubtedly, the baits containing the predators as the protozoa are the biological control option that may be considered for the treatment of the grasshopper breeding sites. The toxic plants are also indicated in controlling these aggressive grasshoppers.

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Explanation of figures

Fig 1: Effect of food plant on live weight (g) of *H. littoralis* hoppers reared on restricted diets of *N. oleander* and *T. alexandrinum* (M±S.E).

Fig 2: Effect of food plant on length (mm) of *H. littoralis* hoppers reared on restricted diets of *N. oleander* and *T. alexandrinum* (M±S.E).



Fig 3: Weight (g) of *H. littoralis* hoppers reared on restricted diet of *T. alexandrinum* or *N. oleander* for 23 days followed by switching diet for seven days (M±S.E).  = nymphs fed on *Trifolium* for 23 days then fed on *Nerium* for 7 days.  = nymphs fed on *Nerium* for 23 days then fed on *Trifolium* for 7 days.



Fig 4: Length (mm) of *H. littoralis* hoppers reared on restricted diet of *T. alexandrinum* or *N. oleander* for 23 days followed by switching diet for seven days (M±S.E).  = nymphs fed on *Trifolium* for 23 days then fed on *Nerium* for 7 days.  = nymphs fed on *Nerium* for 23 days then fed on *Trifolium* for 7 days.

Fig 5: Effect of food plant on the live weight (g) of grasshopper *H. littoralis* female adults reared on either *T. alexandrinum* or *N. oleander*, **Highly significant ($P < 0.01$), ***Very highly significant ($P < 0.001$) between *Trifolium*-fed females and *Nerium*-fed females (M±S.E).

Fig 6: Age specific survivorship of the grasshopper *H. littoralis* cohorts. (A): adults fed *T. alexandrinum*, (B): adults fed *N. oleander*, and (C): adults fed *N. oleander* for 45 days before being switched to *T. alexandrinum*.

Fig 7: Ovary of *H. littoralis* females fed on clover and *Nerium* for 13 days (A & B), ovary of females fed on *Nerium* for 45 days (C), and eggs produced by clover and *Nerium* fed females (D & E). g = germarium; l = lateral oviduct; o = ovary; ov = ovariole.

